Ethosome: a novel vesicular carrier for transdermal drug delivery

Saquib Raza Zahid1, Neeraj Upmanyu1, Surendra Dangi1*, Sudhir Kumar Ray1, Prabhat Jain2, Geeta Parkhe2
1School of Pharmacy & Research Peoples University, Bhopal (M.P.), India
2Scan Research Laboratories, Bhopal (M.P.), India

ABSTRACT

Delivery across skin is striking due to its easy convenience. However, drug delivery across skin is still a confront in biomedical sciences. Over the past few decades, various successful narrative devices and techniques have emerged to optimize drug delivery across skin whose barricading behaviour constricts entry of most of the therapeutic agents. Ethosomes are non-invasive delivery transporter that enables drugs to reach the deep skin layers and/or the systemic circulation. Although ethosomal systems are theoretically sophisticated, they are characterized by simplicity in their preparation, efficacy and safety. A combination that can highly inflate their application. Ethosomes are soft, malleable vesicles adapted for enhanced delivery of active agents. This article reviews work carried out method of preparation, application and characterization of ethosomal systems. Because of their exceptional structure, ethosomes are able to encapsulate and deliver through the skin highly lipophilic molecules such as testosterone, cannabinoids and minoxidil as well as cationic drugs such as trihexyphenidil and propranolol. Results obtained in a double-blind two-armed randomized clinical study showed that treatment with the ethosomal acyclovir formulation appreciably improved all the evaluated parameters. In further work, the ethosomal expertise was broadened to introduce agents into cultured cells and microorganisms. Enhanced delivery of bioactive molecules through the skin and cellular membranes by means of an ethosomal transporter opens numerous confronts and prospects for the research and future development of novel improved therapies.

Keywords: Ethosomes, Skin layers, Characterization

INTRODUCTION

The impending benefits related to transdermal drug delivery are widely reported in the literature. Among these advantages are compact side effects and prevention of first-pass hepatic elimination and intestinal degradation. However, percutaneous delivery of most molecules is often excluded because of the barrier nature of the skin, which functionally impedes the entrance of exogenous materials. To overcome this obstruction, various methods of skin permeation enhancement described in numerous scientific works and patents have been used. Human skin has a multifunctional role including its major role as a barrier against both the outlet of endogenous substances (water) and the entrance of xenobiotic material (chemicals and drugs). It is regarded as first line of protection in human body. The stratum corneum consisting corneocytes mainly accounts for the barrier properties of skin. From some time several aspect have come out which has given immense esteem and rapid progress to transdermal delivery formulations over conventional formulations because circumvention of variations which appear at gastro-intestinal absorption, improvement in bioavailability of drugs by delivering the active principles directly into the systemic circulation, bypassing the hepatic metabolism, by giving a constant, controlled drug enter decreasing the variations in drug plasma levels, augmentation in patient compliance by providing a simplified way of administration, lowest risk of trauma or any other injury of tissue. In order to increase the permeability of the skin for transdermal delivery of drugs several passive as well as active procedure have been proposed such as penetration enhancers, vesicles, iontophoresis, supersaturated systems, electroporation, phophoresis, use of microneedles and jet injectors, etc. Although all the efforts devoted to penetration enhancement, only few bioactive agents are currently transdermally administered. One of the most utilizable methods for drugs’ transport across the skin is the use of...
vesicle formulation as skin delivery systems. Ethosomes are soft, malleable lipid vesicles composed mainly of phospholipids, alcohol in relatively high concentration (20-45%) and water (Fig. 1). Ethosomes were first developed by Touitou and her colleagues in 1997. This transporter presents interesting features correlated with its ability to permeate whole through the human skin due to its high deformability. The physicochemical characteristics of ethosomes allow these vesicular phospholipids as the vesicle forming component of ethosomal system. Phospholipids with various chemical structures like phosphatidyl choline, phosphatidyl ethanolamine are used at concentrations ranging from 0.5-10%.

**Figure 1: Structure of ethosomes**

**Structure of skin**

Stratum corneum is the remotest layer of the epidermis. It consists of 10 to 25 layers of dead, elongated, fully keratinized corneocytes, which are fixed in a matrix of lipid bilayers. It has been exposed that the stratum corneum is the main barrier to penetration through the skin. When a topical formulation is placed on the skin, the active drug is required to penetrate through the stratum corneum into the viable tissue (Fig 2). The limiting factor for these processes is the slow diffusion through the dead horny layer of skin. Stratum corneum behaves as a hydrophobic membrane. The rates of permeation of skin by low and high molecular weight organic non-electrolytes are mostly determined within the stratum corneum.

**COMPOSITION OF ETHOSOMES**

Ethosomes exhibit lipid bilayer like liposomes; however they differ from liposomes in terms of composition (high content of ethanol). The ethosomes are composed of hydroalcoholic or hydro/glycolipid phospholipid in which the concentration of alcohol is relatively high. Ethosomes may contain phospholipids with various chemical structures like phosphatidylcholine, phosphatidic acid, phosphatidylerine, Phosphatidyl ethanolamine, phosphatidyl glycerol, phosphatidyl inositol, alcohol (ethanol or isopropyl alcohol), water and propylene glycol (or other glycols). Some preferred phospholipids such as Phospholipon 90 (PL-90). It is usually employed in a range of 0.5-10% w/w. Cholesterol at concentrations ranging between 0.1-1% can also be added to the preparation. Alcohol, such ethanol and isopropyl alcohol; among glycols, propylene glycol and Transcutol are generally used which may range from 20 to 50% in the final product. In addition to non-ionic surfactants (PEG-alkyl ethers) and cationic lipids (cocoamide, POE alkyl amines, dodecylamine, cetrimide etc) can be combined with the phospholipids in the preparations. The concentration of the non-aqueous phase (alcohol and glycol combination) may range between 22 to 70%. Various additives which are used for formulation of ethosomes are listed in the Table 1.
**Table 1: Different additive employed in formulation of ethosomes**

<table>
<thead>
<tr>
<th>Class</th>
<th>Example</th>
<th>Uses</th>
</tr>
</thead>
<tbody>
<tr>
<td>Phospholipids</td>
<td>Soya phosphatidyl choline</td>
<td>It influences on the size, entrapment efficacy, zeta potential and penetration properties of the vesicles.</td>
</tr>
<tr>
<td></td>
<td>Egg phosphatidyl choline</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Dipalmityl phosphatidyl choline</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Distearyl phosphatidyl choline</td>
<td></td>
</tr>
<tr>
<td>Alcohol</td>
<td>Ethanol</td>
<td>For providing the softness for vesicle membrane</td>
</tr>
<tr>
<td></td>
<td>Isopropyl alcohol</td>
<td>As a penetration enhancer</td>
</tr>
<tr>
<td>Polyglycol</td>
<td>Propylene glycol, Transcutol RTM</td>
<td>As a skin penetration enhancer</td>
</tr>
<tr>
<td>Cholesterol</td>
<td>Cholesterol</td>
<td>For providing the stability to vesicle membrane</td>
</tr>
<tr>
<td>Edge activators</td>
<td>N-DMSO , Tween[22], Span</td>
<td>Enhances skin permeability</td>
</tr>
<tr>
<td>Dye</td>
<td>Rhodamine-123</td>
<td>For characterization study</td>
</tr>
<tr>
<td></td>
<td>Rhodamine red Fluorescence</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Isothiocynate(FITC)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>6 – Carboxy fluorescence</td>
<td></td>
</tr>
<tr>
<td>others</td>
<td>Dicetyl phosphate</td>
<td>Prevent aggregation of vesicles</td>
</tr>
<tr>
<td>Vehicle</td>
<td>Carbopol D-934, HPMC</td>
<td>As a gel former</td>
</tr>
</tbody>
</table>

**Advantage of ethosomal drug delivery system**

In comparison to other transdermal & dermal delivery systems, Ethosomal drug delivery systems contain several advantages. Few advantages are;

1. It contains non-toxic raw material in formulation.
2. Delivery of large molecules (peptides, protein molecules) is possible.
3. Enhanced permeation of drug through skin for transdermal drug delivery.
4. High patient compliance: The ethosomal drug is administered in semisolid form (gel or cream) hence producing high patient compliance.
5. Simple method for drug delivery in comparison to iontophoresis and Phonophoresis and other complicated methods.
6. Ethosomal drug delivery system can be applied widely in Pharmaceutical, Veterinary, Cosmetic fields.
7. Ethosomal system acts as delivery system for a fluorescent probe (quantum dots) to the skin, in terms of quantity and depth.
8. Ethosomes show highest transdermal flux enhances the permeation of drug through deeper layers of skin.
9. Due to intense research toxicological profiles of the ethosome components are well-evaluated and documented in the scientific literature thus the ethosome technology has no large-scale drug development risk.
10. The ethosomal system is passive, non-invasive and is available for immediate commercialization.
11. Drugs entrapped in ethosome having different physicochemical characteristics and olecular sizes are showing high degree of permeation compare to other nano-carriers.
12. Ethosomes improve skin delivery under occlusive and non-occlusive conditions.
13. High market attractiveness for products with proprietary technology. Relatively simple to manufacture with no complicated technical investments required for production of Ethosomes.

**Disadvantages of ethosomal drug delivery**

1. Ethosomes with poor shells may clump together and leads to precipitation.
2. Adequate solubility of the drug in both lipophilic and aqueous environments to reach dermal microcirculation and gain access to the systemic circulation.
3. Skin irritation or dermatitis due to excipients and enhancers of drug delivery systems.
4. Ethosomal administration is not a means to achieve rapid bolus type drug input, rather it usually designed to offer slow, sustained drug delivery.
5. Drugs that require high blood levels cannot be administered –limited to only potent drugs (daily dose ≤10 mg or less).
6. Poor practical yield.
7. Transfer of ethosomes from organic to aqueous layer leads to loss of product.
8. The molecular size of the drug should be reasonable that it should be absorbed percutaneously.
9. Adhesive may not adhere well to all types of skin.
10. May not be economical.
11. In case if shell locking is ineffective then the ethosomes may coalesce and fall apart on transfer into water.

**MECHANISM OF DRUG PENETRATION**

The mechanism of penetration of the ethosomes involves two concurrent mechanisms of ethanol effect and ethosome effect on the stratum corneum lipid bilayer. Because of the use of ethanol in the preparation of the ethosomes, the deformability of the vesicles is increased. The high alcohol content is expected to partially extract the stratum corneum lipids (Fig 3). These processes are responsible for increasing inter and intracellular permeability of ethosomes. The ultra deformable vesicles can move in the path of the disordered stratum corneum and finally release drug in the deeper layers of the skin.

Ethosomes effect

Increased cell membrane lipid fluidity caused by the ethanol of ethosomes results increased skin permeability. So the ethosomes permeates very easily inside the deep skin layers, where it got fused with skin lipids and releases the drugs into deep layer of skin.

Ethanol effect

Ethanol acts as a penetration enhancer through the skin. The mechanism of its penetration enhancing effect is well known. Ethanol penetrates into intercellular lipids and increases the fluidity of cell membrane lipids and decrease the density of lipid multilayer of cell membrane.
METHODS OF PREPARATION

Ethosomes can be prepared and formulated by four methods. All methods are sound simple and convenient because no need of complex processes or sophisticated instruments.

**Cold method**

This is the most common method utilized for the preparation of ethosomal formulation. In this method phospholipid, drug and other lipid materials are dissolved in ethanol in a covered vessel at room temperature by vigorous stirring with the use of mixer. Propylene glycol or other polyol is added during stirring. This mixture is heated to 300°C in a water bath. The water heated to 300°C in a separate vessel is added to the mixture, which is then stirred for 5 min in a covered vessel. The vesicle size of ethosomal formulation can be decreased to desire extent using sonication 32 or extrusion 33 method. Finally, the formulation is stored under refrigeration 34.

Figure 3: Mechanism of action of ethosomes for skin delivery 13
Hot method

In this method phospholipid is dispersed in water by heating in a water bath at 40°C until a colloidal solution is obtained. In a separate vessel ethanol and propylene glycol are mixed and heated to 40°C. Once both mixtures reach 40°C, the organic phase is added to the aqueous one. The drug is dissolved in water or ethanol depending on its hydrophilic/ hydrophobic properties. The vesicle size of ethosomal formulation can be decreased to the desired extent using probe sonication or extrusion method.

Classic method

The phospholipid and drug are dissolved in ethanol and heated to 30°C±1°C in a water bath. Double distilled water is added in a fine stream to the lipid mixture, with constant stirring at 700rpm, in a closed vessel. The resulting vesicle suspension is homogenized by passing through a polycarbonate membrane using a hand extruder for three cycles.

Mechanical dispersion method

Soya phosphotidylcholine is dissolved in a mixture of chloroform: methanol in round bottom flask (RBF). The organic solvents are removed using rotary vacuum evaporator above lipid transition temperature to form a thin lipid film on wall of the RBF. Finally, traces of solvent mixture are removed from the deposited lipid film by leaving the contents under vacuum overnight. Hydration is done with different concentration of hydroethanolic mixture containing drug by rotating the RBF at suitable temperature.

CHARACTERIZATION OF ETHOSOMES

Vesicle size and Zeta potential

Particle size and zeta potential can be determined by dynamic light scattering (DLS) using a computerized inspection system and photon correlation spectroscopy (PCS).

Vesicle shape

Visualization of ethosomes can be done using transmission electron microscopy (TEM) and by scanning electron microscopy (SEM). Visualization by electron microscopy reveals a thosomal formulation exhibited vesicular structure 300-400 nm in diameter. The vesicles seem to be alleable as evident by their imperfect round shape.

Entrapment efficiency

The entrapment efficiency of drug in ethosomes can be measured by the ultracentrifugation technique. The chemical nature of the lipid is an important factor in determining the EE of drug in the ethosomes because lipid which forms bilayer structure holds the drug perfectly. On the other hand, the imperfection of the lipid structure could offer space to accommodate the drug. The vesicles are separated in a high speed cooling centrifuge at 20,000 rpm for 90 minutes in the temperature maintained at 4°C. Separate the sediment and supernatant liquids determine the amount of drug in the sediment by lysing the vesicles using methanol.

\[
\% \text{ Entrapment} = \frac{\text{Actual content}}{\text{Theoretical content}} \times 100
\]

Surface tension measurement

The surface tension activity of drug in aqueous solution can be measured by the ring method in a Du Nouy ring tensiometer.

Surface morphology study

Different types of lipids influence the surface morphology or shape of the particles. Lipid microparticle suspensions were deposited on metallic stubs then placed in liquid nitrogen and dried under vacuum. The freeze-dried microparticles were coated uniformly with gold. It is characterized for morphology and surface properties using a scanning electron microscope.

Drug content

Drug content of the ethosomes can be determined using UV spectrophotometer. This can also be quantified by a modified high performance liquid chromatographic method.

Transition temperature

The Transition temperature (T) of vesicular lipids is measured in duplicate by DSC in an aluminum pan at a heating rate of 10°C per min within a temperature range from 20º-300ºC, under a constant nitrogen stream.

Stability studies

The ability of ethosomal preparations to retain the drug (i.e., drug-retentive behavior) can be checked by keeping the preparations at different temperatures, i.e., 25 ± 2°C (room temperature, RT), 37 ± 2°C and 45 ± 2°C for different periods of time (1, 20, 40, 60, 80 and 120 days). The ethosomal preparations were kept in sealed vials (10 ml capacity) after flushing with nitrogen. The stability of ethosomes was also determined quantitatively by monitoring size and morphology of the vesicles using DLS and TEM.

Degree of deformability and turbidity

The degree of deformability of the Ethosomal preparation can be performed by extrusion method and the turbidity of the preparation can be performed by using nephelometer.
Skin permeation studies

Confocal laser scanning microscopy (CLSM) method is used to determine the depth of penetration from Ethosomes. The ethosomes shows significantly higher skin deposition possibly due to combined effect of ethanol and phospholipid thus providing a mode for dermal and transdermal delivery.

In vitro drug release study and Drug deposition study

In vitro drug release study and drug deposition of ethosomal preparation can be performed by Franz diffusion cell with artificial or biological membrane, Dialysis bag diffusion.

Phospholipid-ethanol interaction

The Phospholipid-ethanol interaction was studied by using Proton decoupled 31P-NMR and Differential Scanning calorimetry.

EVALUATION OF ETHOSOME 38

Vesicle-skin interaction study by TEM and SEM

From animals ultra-thin sections were cut (Ultracut, Vienna, Austria), collected on formvar coated grids and examined under transmission electron microscope. For SEM analysis, the sections of skin after dehydration were mounted on stubs using an adhesive tape and were coated with gold palladium alloy using a fine coat ion sputter coater. The sections were examined under scanning electron microscope.

Filter membrane-vesicle interaction study by scanning electron microscopy

Vesicle suspension (0.2 mL) was applied to filter membrane having a pore size of 50 nm and placed in diffusion cells. The upper side of the filter was exposed to the air, whereas the lower side was in contact with PBS (phosphate buffer saline solution), (pH 6.5). The filters were removed after 1 hour and prepared for TEM studies by fixation at 4°C in Karnovsky’s fixative overnight followed by dehydration with graded ethanol solutions (30%, 50%, 70%, 90%, 95%, and 100% vol/vol in water). Finally, filters were coated with gold and examined in SEM.

Vesicle-skin interaction study by fluorescence microscopy

Fluorescence microscopy was carried according to the protocol used for TEM and SEM study. Paraffin blocks are used, were made, 5-μm thick sections were cut using microtome (Erma optical works, Tokyo, Japan) and examined under a fluorescence micro Cytotoxicity Assay MT-2 cells (Tlymphoid cell lines) were propagated in Dulbecco’s modified Eagle medium (HIMEDIA, Mumbai, India) containing 10% fetal calf serum, 100 U/mL penicillin, 100 mg/mL streptomycin, and 2 mmol/L L-glutamine at 37°C under a 5% CO2 atmosphere. Cytotoxicity was expressed as the ratio of the cytotoxic dose 50 (CD50) that induced a 50% reduction of absorbance at 540nm.

Skin permeation studies

The hair of test animals (rats) were carefully trimmed short (<2 mm) with a pair of scissors, and the abdominal skin was separated from the underlying connective tissue with a scalpel. The excised skin was placed on aluminum foil, and the dermal side of the skin was gently teased off for any adhering fat and/or subcutaneous tissue. The effective permeation area of the diffusion cell and receptor cell volume was 1.0 cm2 and 10 mL, respectively. The temperature was maintained at 32°C ± 1°C. The receptor compartment contained phosphate buffer saline solution (10 mL of pH 6.3). Excised skin was mounted between the donor and the receptor compartment. Ethosomal formulation (1.0 mL) was applied to the epidermal surface of skin. Samples (0.5 mL) were withdrawn through the sampling port of the diffusion cell at 1, 2, 4, 8, 12, 16, 20 & 24 hour time intervals and analyzed by high performance liquid chromatography assay.

HPLC Assay

The amount of drug permeated in the receptor compartment during in vitro skin permeation experiments and in MT-2 cell was determined by HPLC assay.

Drug uptake studies

The uptake of drug into MT-2 cells (1×106 cells/mL) was performed in 24-well plates (Corning Inc) in which 100 μL RPMI medium was added. Cells were incubated with 100 μL of the drug solution in phosphate buffer saline solution (pH 7.4), ethosomal formulation, or marketed formulation, and then drug uptake was determined by analyzing the drug content by HPLC assay.

Statistical analysis

Statistical significance of all the data generated was tested by employing ANOVA followed by studentized range test. A confidence limit of P < .05 was fixed for interpretation of the results using the software PRISM (GraphPad, Version 2.01, San Diego, CA).

APPLICATION OF ETHOSOMES

Ethosomes find its diverse applications in various categories of drugs like Antifungal, Antibiotics, Skin infections and Cosmetic field. Some are listed as below (Table 2).

<table>
<thead>
<tr>
<th>Principle ingredients</th>
<th>Formulation</th>
<th>Rationale of ethosomal delivery</th>
<th>Application</th>
</tr>
</thead>
<tbody>
<tr>
<td>5-aminolevulivic acid(ALA)</td>
<td>5-aminolevulivic Acid ethosomes</td>
<td>Significantly improved the delivery of ALA in the inflammatory skin.</td>
<td>Anti- psoriasis</td>
</tr>
<tr>
<td>Erythromycin</td>
<td>Erythromycin ethosomes</td>
<td>Ethosomal erythromycin was highly efficient in eradicating S. aureus- induced intradermal infections</td>
<td>Anti bacterial</td>
</tr>
<tr>
<td>Isoeugenol</td>
<td>Isoeugenol ethosomes</td>
<td>Chemicals (allergen) in vesicular carrier system can enhance the sensitizing capacity.</td>
<td>Allergen</td>
</tr>
<tr>
<td>Matrine</td>
<td>Matrine ethosomes</td>
<td>Improves the percutaneous permeation</td>
<td>Anti- inflammatory</td>
</tr>
<tr>
<td>Methotrexate</td>
<td>Methotrexate ethosomes</td>
<td>Ethosomes showed favourable skin permeation characteristic</td>
<td>Anticancer</td>
</tr>
<tr>
<td>Minoxidil</td>
<td>Minoxidil ethosomes</td>
<td>Enhance the penetration and accumulation of minoxidil in the skin by Pilosebaceous targeting</td>
<td>Hair growth promoter</td>
</tr>
</tbody>
</table>
ent past.

This delivery of

alh

offers a good opportunity for the non-physician to make the therapy more effective. Ethosomes allow better control over drug release agents more effective. Further, research in this area will ethosomes in drug delivery. Different reports show a promising future of

initiated a new area in vesicular research for transdermal

penetration of drugs lipid vehicle based enhancement

main barrier layer for penetration of drug. Various

For transdermal delivery of drugs, stratum corneum is the

Body

Skin genuity

Noicellex

Decorin cream

Cellutight EF

In 2000, the ethosomes technology began to Commercialize. There are many companies which developed ethosomes

Marketed product of ethosomes

In 2000, the ethosomes technology began to Commercialize. There are many companies which developed ethosomes products (Table 3).

<table>
<thead>
<tr>
<th>Name of product</th>
<th>Uses</th>
<th>Manufacturer</th>
</tr>
</thead>
<tbody>
<tr>
<td>Testosterone ethosomes</td>
<td>Testosterone ethosomes for Enhanced transdermal delivery</td>
<td>Steroid hormone</td>
</tr>
<tr>
<td>Trihexyphenidyl HCL ethosomes</td>
<td>Increased drug entrapment efficiency, reduced side effect and constant systemic levels</td>
<td>Anti-parkinsonian</td>
</tr>
<tr>
<td>Acyclovir ethosomes</td>
<td>Binary combination of the Lipophilic drug ACV-C16 and the ethosomal ACV absorption into synergistically the skin</td>
<td>Anti-viral</td>
</tr>
<tr>
<td>Azelaic acid ethosomes</td>
<td>Release rate was higher from ethosomes than from liposomes</td>
<td>Anti- keratinizing</td>
</tr>
<tr>
<td>Bacitracin ethosomes</td>
<td>Ethosomal enhances intracellular delivery of and reduced drug toxicity in the skin</td>
<td>Polypeptide antibacterial</td>
</tr>
<tr>
<td>Colchicine ethosomes</td>
<td>Enhance skin accumulation, prolong release and improve the specificity</td>
<td>Anti-gout</td>
</tr>
<tr>
<td>Finasteride ethosomes</td>
<td>Enhanced percutaneous absorption of finasteride 5-a reductase inhibitor</td>
<td>Anti-Fungal</td>
</tr>
<tr>
<td>Fluconazole ethosomes</td>
<td>enhances the skin permeation</td>
<td>Anti-Fungal</td>
</tr>
<tr>
<td>Ibuprofen ethosomes</td>
<td>Transdermal nanosystem, designed by using an ethosomal carrier</td>
<td>Antipyretic</td>
</tr>
<tr>
<td>Ligustrazine ethosomes</td>
<td>Ethosome patch enhances the permeation the skin</td>
<td>Pulmonary vasodilator</td>
</tr>
<tr>
<td>Salbutamol ethosomes</td>
<td>Enhanced drug delivery through skin with ethosomes</td>
<td>Anti-asthmatic</td>
</tr>
<tr>
<td>Sotalol ethosomes</td>
<td>Enhances the systemic absorption</td>
<td>anti-arrhythmic</td>
</tr>
<tr>
<td>Vitamin A, C, E ethosomes</td>
<td>Anti-oxidation of phospholipid was increase due to the synergistic interaction of all three together as compare to individual use</td>
<td>Vitamin</td>
</tr>
<tr>
<td>Stavudine ethosomes</td>
<td>Ethosome increase the transdermal flux, prolong the release of Stavudine</td>
<td>Antiretroviral</td>
</tr>
</tbody>
</table>

Table 3: Marketed formulations of ethosomes

Future perspective

For transdermal delivery of drugs, stratum corneum is the main barrier layer for penetration of drug. Various methods have been discovered to enhanced skin penetration of drugs lipid vehicle based enhancement approach has drawn considerable interest in recent past. Studies will continue further to improve skin delivery of drug using lipid vesicles. Introduction of ethosomes has initiated a new area in vesicular research for transdermal drug delivery. Different reports show a promising future of ethosomes in making transdermal delivery of various agents more effective. Further, research in this area will allow better control over drug release in vivo, allowing physician to make the therapy more effective. Ethosomes offers a good opportunity for the non-invasive delivery of small, medium and large sized drug molecules. The results of the first clinical study of acyclovir-ethosomal formulation support this conclusion. Multititer quantities of ethosomal formulation can be prepared very easily. It, therefore, should be not before long that the corresponding drug formulation would have found their way into clinics to be tested for widespread usage. Thus, it can be a logical conclusion that ethosomal formulations possess promising future in effective dermal/transdermal delivery of bioactive agents.

Patented

Ethosome was invented and patented by Prof. Elka Touitou along with her students of Department of Pharmaceutic at the Hebrew University School of Pharmacy 33, 41.
Novel Therapeutic Technologies Inc (NTT) of Hebrew University has been succeeded in bringing a number of products to the market based on ethosome delivery system Table 4.

### Table 4: Patents claimed for ethosome formulations

<table>
<thead>
<tr>
<th>Title</th>
<th>Inventor</th>
<th>Patent no</th>
<th>Year</th>
<th>Results</th>
</tr>
</thead>
<tbody>
<tr>
<td>Tretinoin ethosomes gel and preparation method thereof</td>
<td>Hu Chunmei, Liu Yan, Wang Jing, Li Rong</td>
<td>CN104983675 A</td>
<td>2015</td>
<td>the prepared tretinoin ethosomes gel is an externally-used transdermal delivery preparation</td>
</tr>
<tr>
<td>Chinese medicinal ethosome gel patch for treating herpes zoster and preparation method</td>
<td>Bu Ping; Hu Rong; Chen Lin; Wei Rong; Wu Huanhuan; Huang Xiaoli</td>
<td>CN103536700 (A)</td>
<td>2014</td>
<td>Easy in medication and convenient to use, has a good therapeutic effect, quick response,</td>
</tr>
<tr>
<td>Ethosome gel film-coating agent with multiple wound repair effects and preparation method of ethosome gel film-coating agent 1</td>
<td>Chen Jie; Huang Changping; Zheng Maoxin; Nie Kaipin</td>
<td>CN103893394 (A)</td>
<td>2014</td>
<td>The Ethosome entrapped film-coating agent helps to promote healing and nutrition supplying of the wound tissue.</td>
</tr>
<tr>
<td>Chinese medicinal ethosome gel patch for treating herpes zoster and preparation method</td>
<td>Bu Ping; Hu Rong; Chen Lin; Wei Rong; Wu Huanhuan; Huang Xiaoli</td>
<td>CN103536700 (A)</td>
<td>2014</td>
<td>Easy in medication and convenient to use, has a good therapeutic effect, quick response,</td>
</tr>
<tr>
<td>Ethosome preparation of male hormone</td>
<td>Shu Meng; Jianxin Li; Yanmin Guan</td>
<td>CN102406605 (A)</td>
<td>2012</td>
<td>To improve transdermal transport of male hormone</td>
</tr>
<tr>
<td>Paclitaxel ethosome gel and preparation method thereof</td>
<td>Jianping Tan; Lixin Jiang; Tanran Chang; Zhiwen Zhou</td>
<td>CN102579323 (A)</td>
<td>2012</td>
<td>The action of stimulation to the skin can be reduced, and the percutaneous permeation effect is good.</td>
</tr>
<tr>
<td>Acyclovir ethosome and preparation method thereof</td>
<td>Xuewen Wu; Yan Xiong</td>
<td>CN102133183 (A)</td>
<td>2011</td>
<td>Acyclovir ethosome has high stability and narrow particle size distribution</td>
</tr>
<tr>
<td>Podophyllotoxin ethosomes and preparation methods thereof</td>
<td>Nianping Feng; Yanyan Yu; Jihui Zhao; Haiting Weng; Xiaoqin Shi</td>
<td>CN102144972 (A)</td>
<td>2011</td>
<td>The invention discloses two preparation methods for the podophyllotoxin ethosomes</td>
</tr>
</tbody>
</table>

**CONCLUSION**

Ethosomes are a pioneering platform technology that offers modified delivery systems to meet the necessary criteria for efficient and safe drug administration into and across the skin. Considerable research has been conducted to characterize this novel non-invasive carrier. Because of their composition, ethosomes are soft vesicles, which penetrate through the SC lipids and transport the active agent to the deep layers of the skin. These vesicular systems were proven efficient for dermal and transdermal delivery of various energetic agents in animals and humans. Among the many areas where ethosomal transdermal delivery could be helpful are Parkinson’s disease, hormone replacement therapy, cardiovascular treatments and rheumatic disorders. Delivery of antibiotics and antiviral to the deep layers of the skin and drugs for management of alopecia to the hair follicles by an ethosomal carrier could greatly increase drug treatment competence and patient convenience. The highly competent delivery, together with its lack of toxicity and preparation simplicity, makes this system a promising candidate for the administration of chemical and biological compounds. Such developments may further increase the field of transdermal applications.
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